

Discovering and validating biological hypotheses from coherent patterns in functional genomics data

Marcin P. Joachimiak

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Environmental Microbiology Sessions

Integrated omics in systems biology: The new frontier for environmental biotechnology

Convener: Terry C. Hazen, Univ. of California, Berkeley, CA

Original: “Transcriptomics and Bioinformatics”

“Discovering and validating biological hypotheses from coherent patterns in functional genomics data”

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Abstract

The area of transcriptomics analysis is among the more established in computational biology, having evolved in both technology and experimental design. Transcriptomics has a strong impetus to develop sophisticated computational methods due to the large amounts of available whole-genome datasets for many species and because of powerful applications in regulatory network reconstruction as well as elucidation and modeling of cellular transcriptional responses. While gene expression microarray data can be noisy and comparisons across experiments challenging, there are a number of sophisticated methods that aid in arriving at statistically and biologically significant conclusions. As such, computational transcriptomics analysis can provide guidance for analysis of results from newer experimental technologies. More recently, search methods have been developed to identify modules of genes, which exhibit coherent expression patterns in only a subset of experimental conditions. The latest advances in these methods allow to integrate multiple data types and datasets, both experimental and computational, within a single statistical framework accounting for data confidence and relevance to specific biological questions. Such frameworks provide a unified environment for the exploration of specific biological hypothesis and for the discovery of coherent data patterns along with the evidence supporting them.

ESPP2 is part of the Virtual Institute for Microbial Stress and Survival supported by the U. S. Department of Energy, Office of Science, Office of Biological and Environmental Research, Genomics Program:GTL through contract DE-AC02-05CH11231 between Lawrence Berkeley National Laboratory and the U. S. Department of Energy.

Large scale biology: towards model cells and organisms

Focused on rapidly inferring as much as possible about a cell or organism:

- its physiology,
- the networks that control its behavior,
- and how the resultant phenotypes allow them to survive in diverse and uncertain environments.

Inference based on collaborative, high throughput experiments and comparative analysis.

Yeast community

Environment Stress Pathway Project (ESPP)

Virtual Institute for Microbial Stress and Survival (VIMSS)

Protein Complex Analysis Project (PCAP)

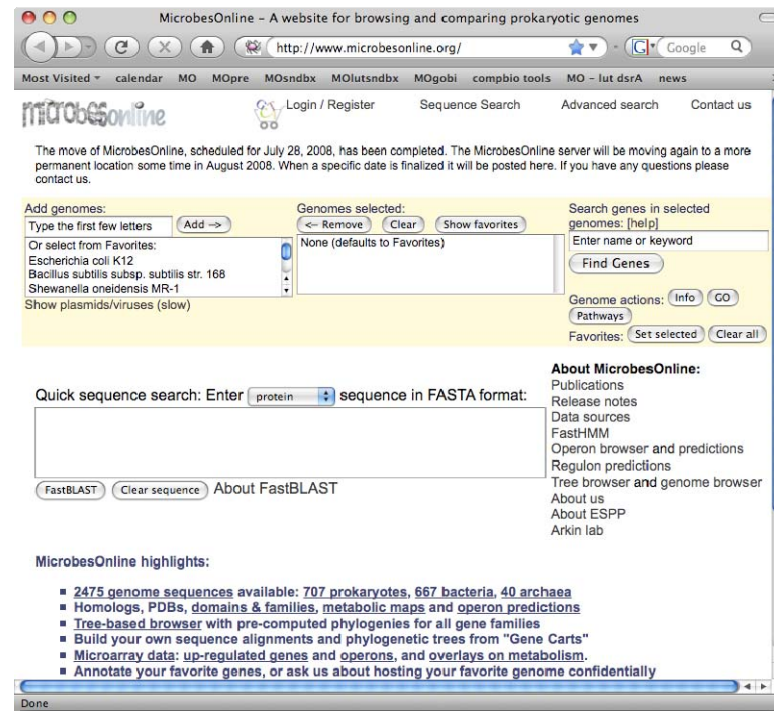
Data from large scale biology projects

- Gene
 - Function
 - Cis-regulatory site
 - Phenotype/fitness (upon gene knockout)
 - Phylogenetic distribution
- Molecular species
 - Relative or absolute level
 - Transcriptomics
 - Proteomics
 - Metabolomics
 - Interaction with other molecular species
 - Protein-protein interactions
 - Transcription factor-DNA binding

Bringing it all together physically and contextually

- Each individual experiment has merit and can be interpreted in isolation.
- However, to achieve the large scale goals it is necessary to integrate datasets and gain multiple contexts.

www.microbesonline.org



Gene function inference

In absence of direct experimental data or characterized close orthologs ...

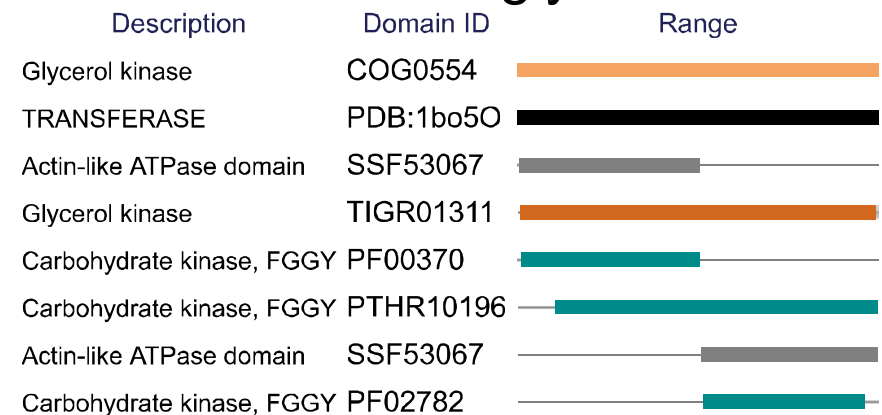
- Sequence similarity
- Domain architecture
- Functional residues
- Genome location context and presence of expected associates
- 3D structure modeling

Legend

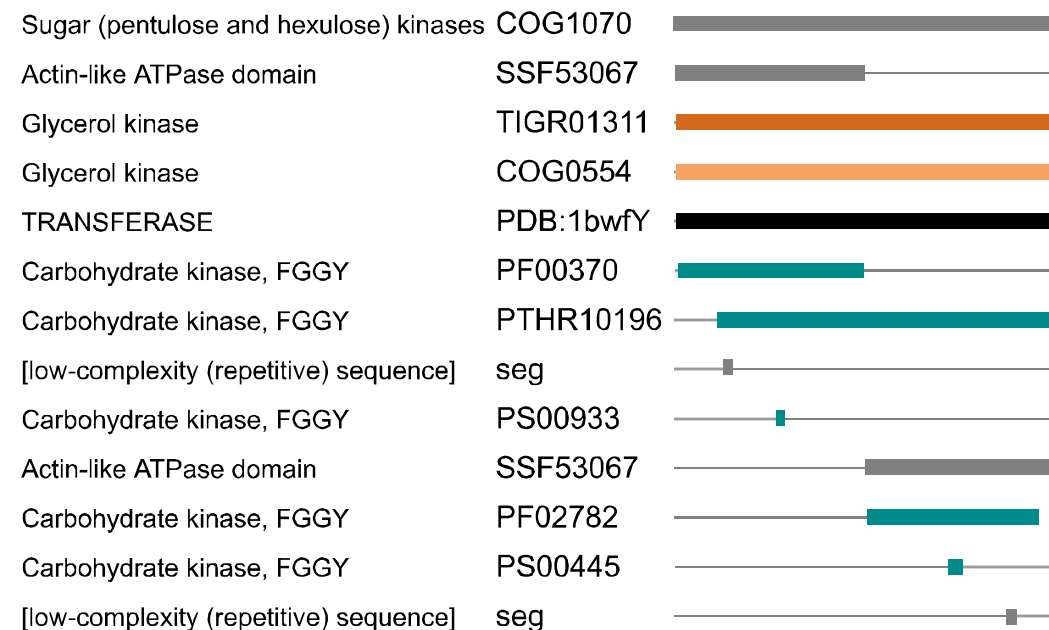
InterPro: IPR000577:  IPR005999: 

Best COG:  No IPR/Other COGs:  PDBs: 

E. coli glycerol kinase

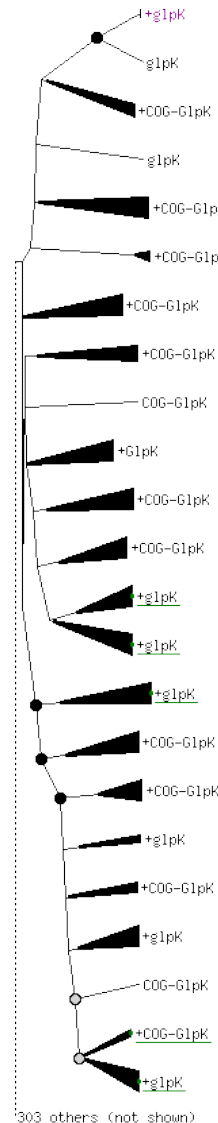


Desulfovibrio vulgaris Hildenborough ortholog



Tree and genome browser for
3,699,361 proteins in
457,623 families from
1076 microbial genomes
(includes multiple family assignments)

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3,699,361 proteins in
457,623 families from
1076 microbial genomes
(includes multiple family assignments)



Genomic map of the *Desulfovibrio vulgaris* Hildenborough (+ strand, 3276764..3286764) and other bacterial genomes. The map shows the arrangement of genes (represented by colored arrows) and their corresponding coordinates (in kb) along the chromosome. The genes are color-coded by function: blue for metabolism, green for information, red for transport, orange for other, and grey for unknown. The map includes the following genes and their coordinates:

- Desulfovibrio vulgaris* Hildenborough (+ strand, 3276764..3286764): *COG-DesR*, *COG-GlpA*, *glpF*, *glpK*, *mdaB*, *COG-NfnB*, *fabG*.
- Desulfovibrio desulfuricans* G20 (- strand, 268790..278790): *COG-DesR*, *COG-GlpA*, *glpF*, *glpK*, *COG3127*, *COG4181*, *iesA*.
- Deinococcus geothermalis* contig (+ strand, 20114..30114): *COG-BaeS*, *COG-DesR*, *COG-GlpA*, *glpF*, *glpK*, *COG-GlpA*, *COG-EutG*, *COG-CalC*.
- Salinibacter ruber* DSM 13855 (- strand, 1829565..1839565): *SRU_1475*, *flp*, *COG-GspA*, *COG-AraC*, *glpK*, *COG-MalK*, *COG-UspA*, *COG-UspE*.
- Acidobacterium bacterium* Ellin345 (+ strand, 3533064..3543064): *COG-CcmA*, *Acid345_2894*, *COG-RstW*, *COG-GlpF*, *COG-GlpK*, *COG-PylG*, *COG-VicK*, *COG-OmpR*, *COG-GleD*.
- Borrelia burgdorferi* B31 (+ strand, 242350..252350): *COG-Lnt*, *BB0238*, *COG1428*, *COG-GlpF*, *COG-GlpK*, *COG-GlpA*, *COG1728*, *COG-CcmAB0246*.
- Symbiobacterium thermophilum* IAM 14863 (- strand, 1320973..1330973): *COG3694*, *COG4587*, *COG4588*, *COG-GdnA*, *COG-GlpK*, *STH1194*, *STH1193*, *COG212*, *COG4770*.
- Microscilla marina* ATCC 23134 contig (+ strand, 118199..128199): *M23134_08114*, *COG3173*, *M23134_08116*, *COG-GlpK*, *COG-GlpA*, *COG-BioF*.
- Halothermothrix orenii* H 168 contig (- strand, 215902..225902): *COG-LeuA*, *COG-LeuB*, *COG-SsdC*, *COG2110*, *COG-GlpK*, *COG579*, *COG-TxeB*, *COG386*, *COG-GlpP*.
- Thermoanaerobacter tengcongensis* (- strand, 1946982..1956982): *TTE2007*, *DnaI_3*, *TTE2005*, *glpF*, *glpF*, *glpK*, *COG579*, *HcaD2*, *AlcC2*.
- Shewanella* sp. PV-4 contig (- strand, 13536..23536): *COG-LeuF*, *COG-LeuD*, *COG-FadL*, *ShewDRA*, *COG-GlpK*, *COG-Fdx*, *ShewDRAFT_0568*, *COG2001*, *COG275*.
- Fusobacterium nucleatum* subsp. *vincentii* ATCC 49256 contig (+ strand, -3881..6118): *COG-GlpA*, *COG-DakK*.
- Pseudomonas aeruginosa* PAO1 (+ strand, 4009444..4019444): *COG3384*, *COG-GlpK*, *COG-EldC*, *glpF*, *glpK*, *glpR*, *glpD*, *COG-MhpC*.
- Escherichia coli* O157:H7 EDL933 (- strand, 4980296..4990296): *menK*, *mmgG*, *Z5475*, *Z5474*, *glpF*, *glpK*, *glpX*, *fpr*, *yiiT*, *yilR*, *yilQ*.
- Enterococcus faecalis* V583 (- strand, 1866976..1876976): *COG-HcaD*, *EF1931*, *glpK*, *COG-GlpA*, *glpF*, *COG-XnaCEFF1925*.
- Stenotrophomonas maltophilia* R551-3 contig (+ strand, 2966..12966): *COG-AceF*, *COG-MutA*, *COG-OmpW*, *COG-GlpA*, *COG-GlpK*, *COG-NlpA*, *COG2378*, *COG3137*, *COG-HemC*.
- Staphylococcus saprophyticus* (- strand, 1506643..1516643): *COG-MutC*, *COG-GlpF*, *COG-Lp2A*, *glpF*, *glpK*, *COG-GlpA*, *COG-PidB*, *COG-MiaA*.
- Bacillus clausii* KSM-K16 (+ strand, 3506775..3516775): *COG-Act2*, *COG-PurR*, *glpF*, *glpK*, *COG-FabG*, *COG2378*, *COG-SodC*, *folD*.
- Exiguobacterium sibiricum* 255-15 contig (- strand, 5176..15176): *COG-PmrR*, *COG-PmrR*, *COG-LeuA*, *COG2380*, *COG-GlpF*, *COG-GlpK*, *COG-DAK1*, *COG-DAK1*, *COG-AziC*.
- Bacillus halodurans* C-125 (+ strand, 1173986..1183986): *BH1089*, *BH1090*, *glpF*, *glpK*, *BH1094*, *glpD*, *BH1096*.
- Exiguobacterium sibiricum* 255-15 contig (- strand, 221756..231756): *COG-CcmA*, *ExpDRAFT_171003264*, *COG-GlpF*, *COG-GlpF*, *COG-GlpK*, *COG-GlpA*, *COG-FmE*, *COG-LpA*.
- Geobacillus kaustophilus* HTA426 (+ strand, 1374806..1384806): *COG3344*, *SK1358*, *COG-GlpF*, *COG-GlpK*, *COG2199*, *COG-GloACOG-Paol*, *COG-Paol*.
- Bacillus subtilis* subsp. *subtilis* str. 168 (+ strand, 998422..1008422): *vhcX*, *vhcA*, *glpF*, *glpD*, *vhcB*, *vhcY*.



- RegTransBase provides information on microbial transcription factor binding sites and sequence motifs based on expert curation and literature.

Articles Curated: 4,445

Experiments: 10,216

Organisms: 180

Genes: 17,346

Sites: 8,833

Regulators: 971

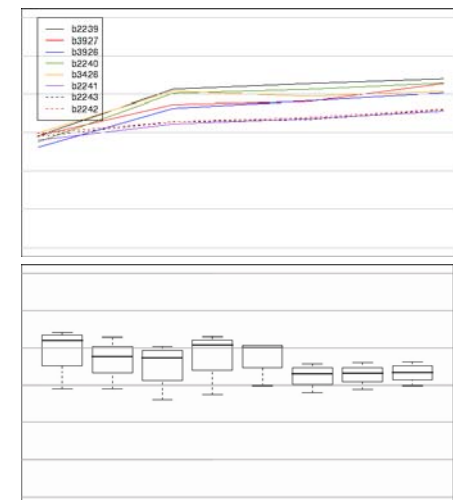
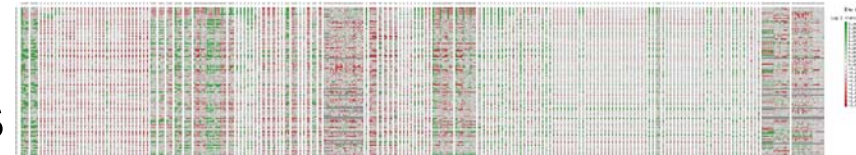
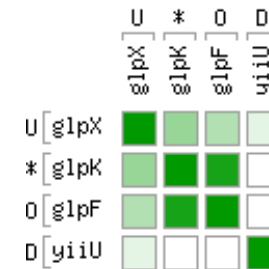
Effectors: 794

- The two databases and websites are interlinked with ongoing development efforts for new functionalities.

MicrobesOnline data analysis

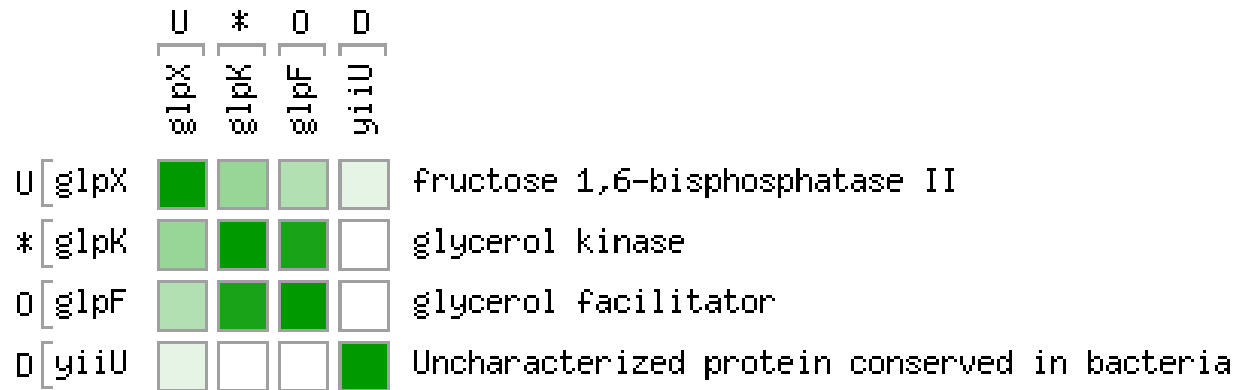
Currently focused on gene expression data but generalizable to many data types.

- Gene-gene and experiment-experiment correlations *
- Gene expression profile searches (functional profiling)
- Line and box plots allowing to subset on genes and experiments *



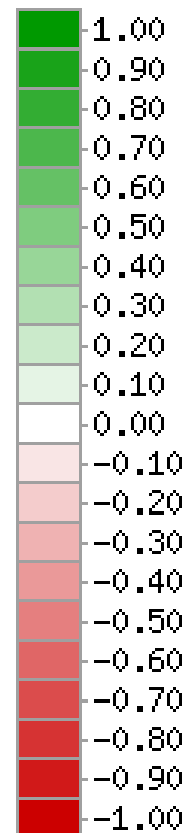
* Upcoming release of
www.microbesonline.org

Analysis: gene-gene expression correlations



No Data

Pearson correlation coefficient



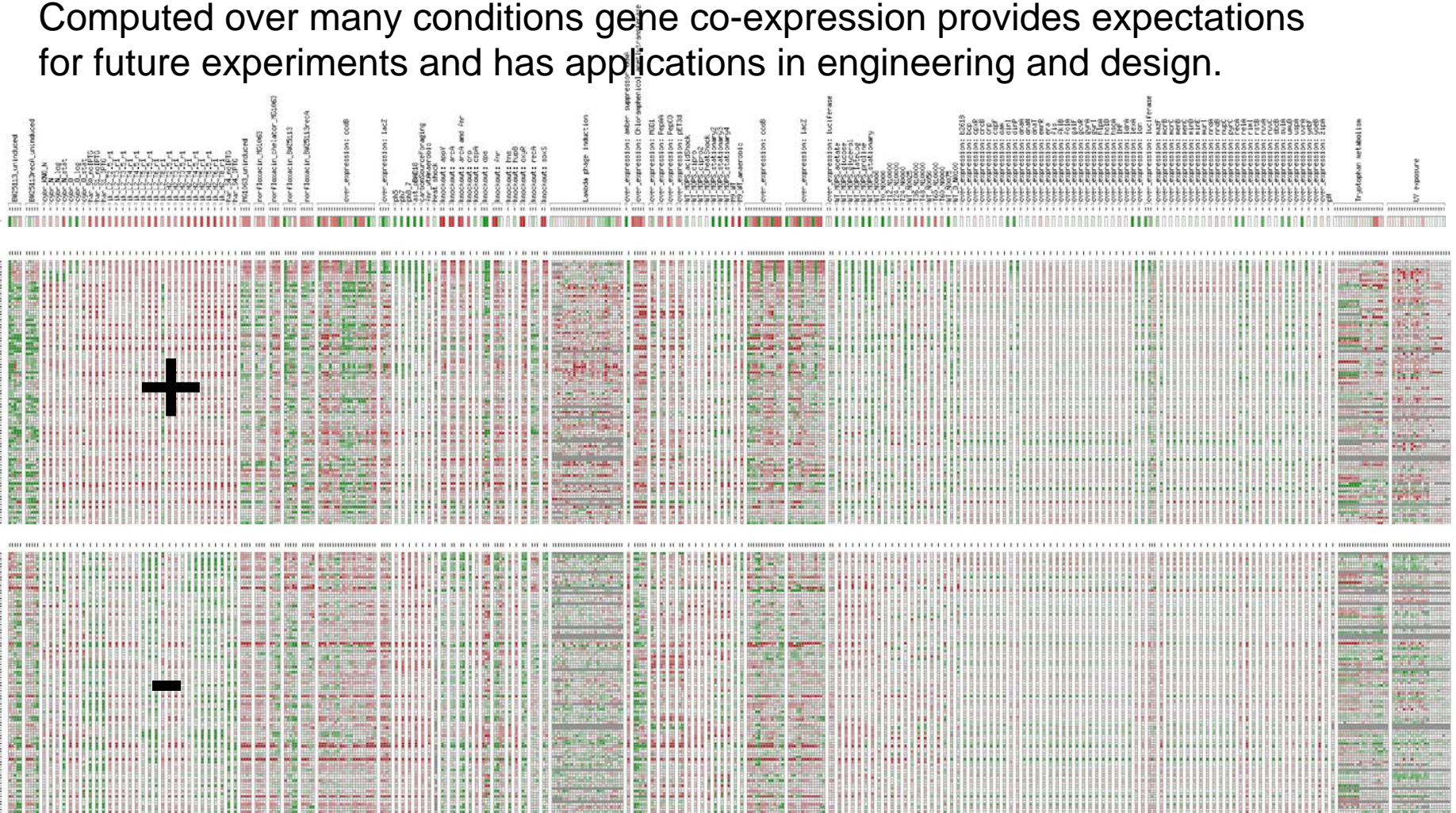
Summarizes similarity in gene expression for a set of genes across a set of experiments.

Genes flanking an operon provide a context for significance as well as operon assignments.

Average correlations of randomly sampled genes or permuted gene expression data for the genes of interest can provide statistical significance.

Gene expression profile searches

Searches rely on the Pearson correlation coefficient as the similarity measure. Can identify genes with similar as well as opposite expression patterns. Computed over many conditions gene co-expression provides expectations for future experiments and has applications in engineering and design.



Identifying candidate genes with gene expression profile searches

This example query profile was based on the mean expression of two *E. coli* genes from a single operon. A biologically motivated cutoff appeared based on a group of clearly functionally related genes from the glycerophospholipid metabolism pathway.

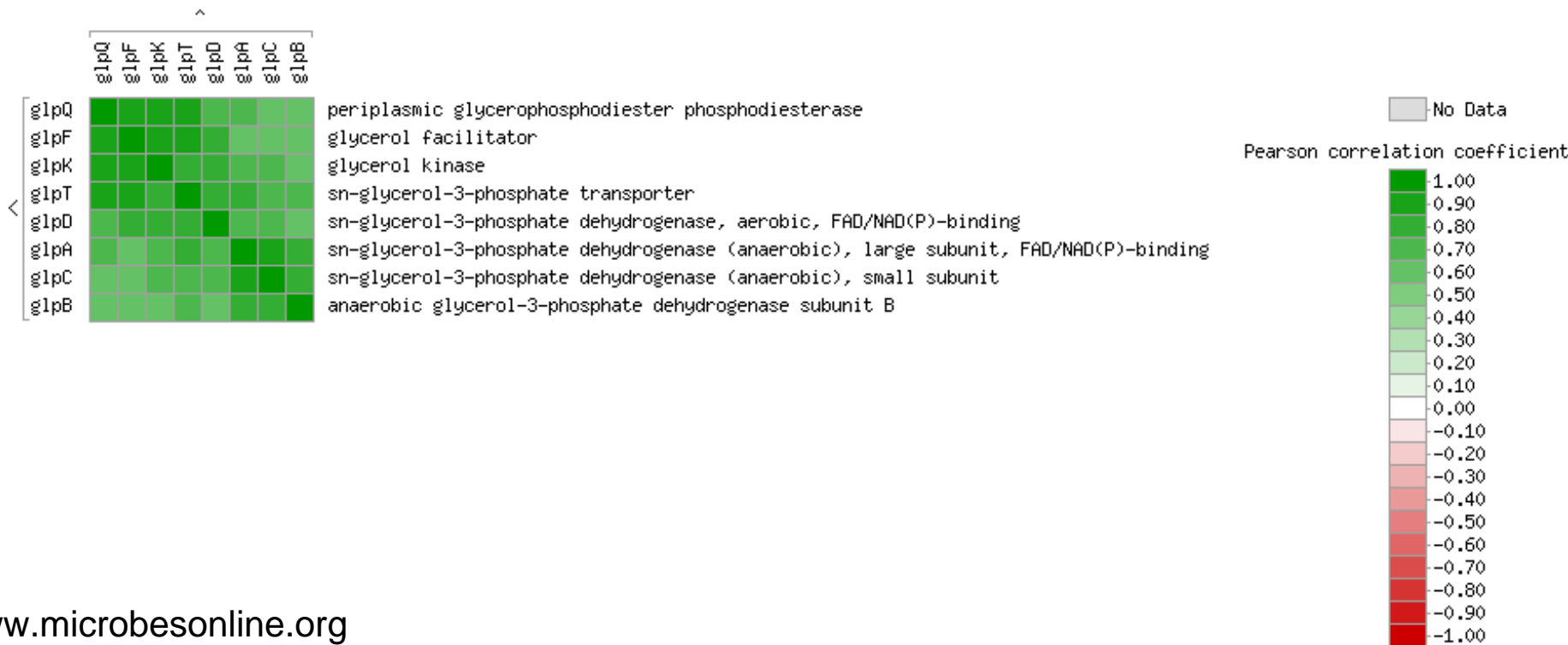


Correlation Coefficient	Number of Data Points	Gene Id	Gene Description
0.98	317	glpF*	glycerol facilitator
0.97	311	glpK*	glycerol kinase
0.88	317	glpQ	periplasmic glycerophosphodiester phosphodiesterase
0.87	316	glpT	sn-glycerol-3-phosphate transporter
0.77	313	glpD	sn-glycerol-3-phosphate dehydrogenase, aerobic, FAD/NAD(P)-binding
0.69	317	glpA	sn-glycerol-3-phosphate dehydrogenase (anaerobic), large subunit, FAD/NAD(P)-binding
0.65	316	glpC	sn-glycerol-3-phosphate dehydrogenase (anaerobic), small subunit
0.60	317	glpB	anaerobic glycerol-3-phosphate dehydrogenase subunit B
0.45	316	mglA	fused methyl-galactoside transporter subunits of ABC superfamily: ATP-binding components

Analysis of candidates: gene-gene expression correlations

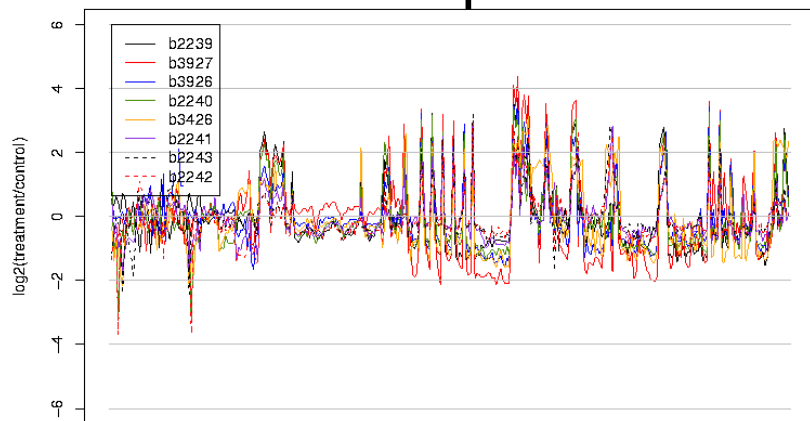
Can expand analysis to genome-wide with clustering, but standard clustering is limited and eventually need more sophisticated and computationally intensive bicluster search methods.

For example, need the ability to seed a search with specific genes and experiments.

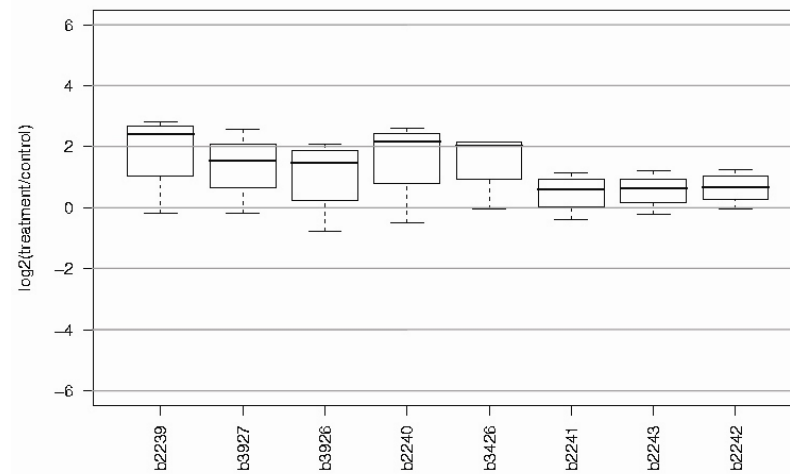
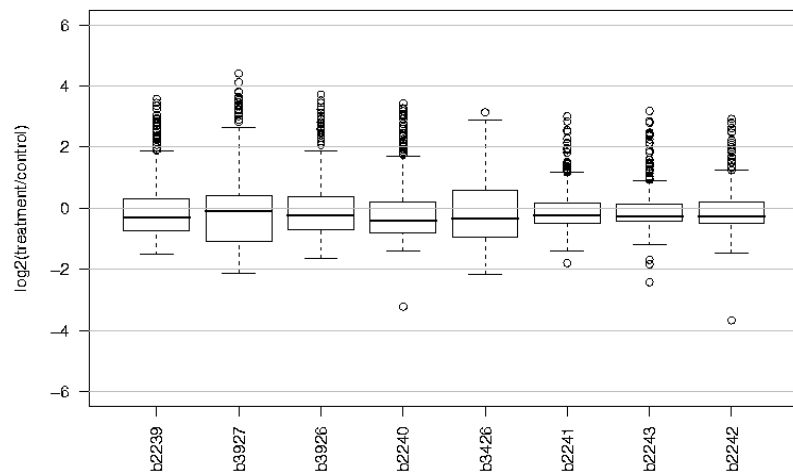
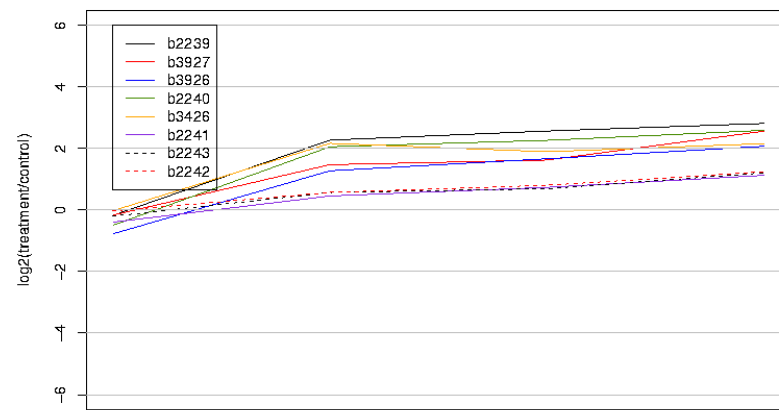


Analysis of candidates: line and box plots

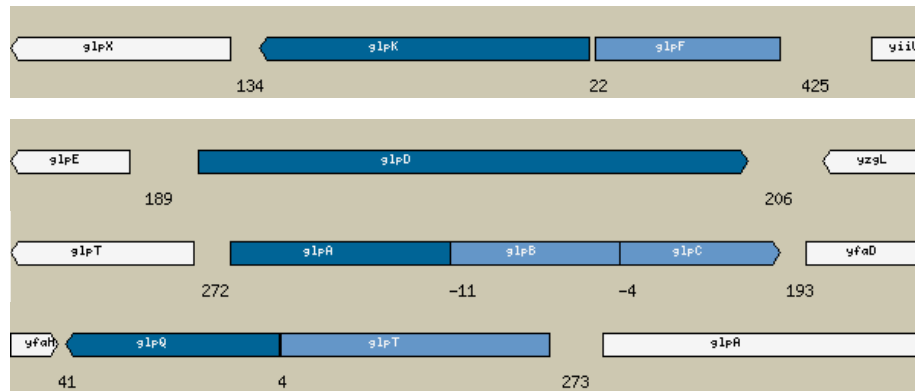
Entire compendium



pH 2, 5, 7, 8.7



Analysis of candidates: iterative profile searches



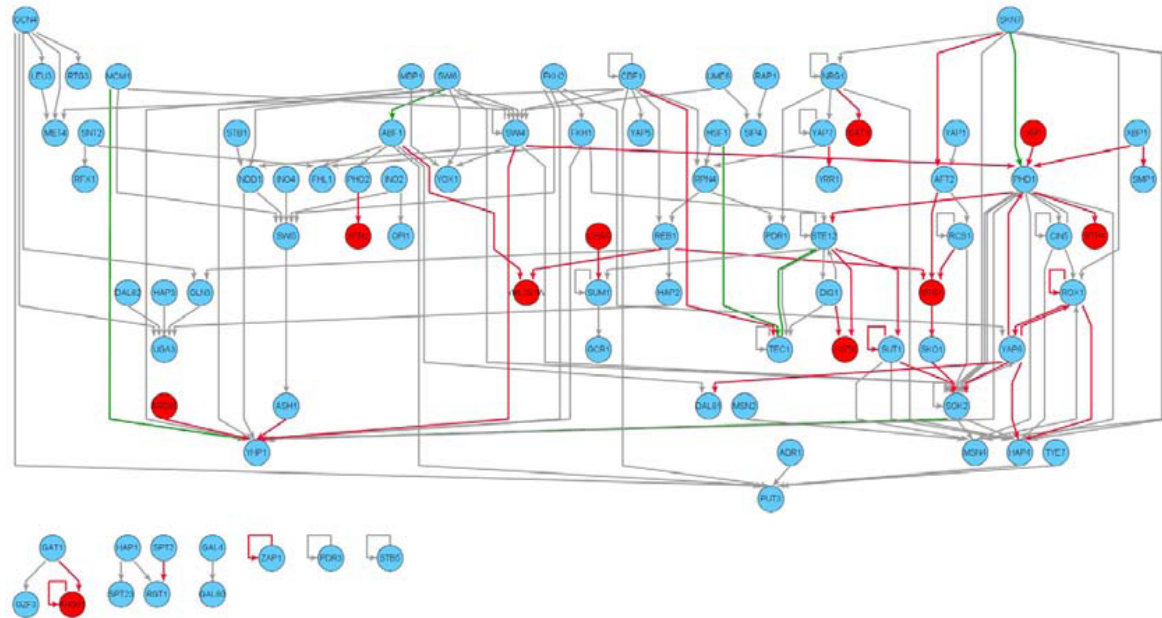
Six additional genes (from 3 different operons) were identified with high expression profile similarity to the query. A natural cutoff was provided by the next best weakly correlated gene. All top hits are members of the glycerophospholipid metabolism pathway.

Correlation Coefficient	Number of Data Points	Gene Id	Gene Description
0.93	316	glpT*	sn-glycerol-3-phosphate transporter
0.93	311	glpK*	glycerol kinase
0.91	317	glpF*	glycerol facilitator
0.91	317	glpQ*	periplasmic glycerophosphodiester phosphodiesterase
0.86	313	glpD*	sn-glycerol-3-phosphate dehydrogenase, aerobic, FAD/NAD(P)-binding
0.85	317	glpA*	sn-glycerol-3-phosphate dehydrogenase (anaerobic), large subunit, FAD/NAD(P)-binding
0.82	316	glpC*	sn-glycerol-3-phosphate dehydrogenase (anaerobic), small subunit
0.79	317	glpB*	anaerobic glycerol-3-phosphate dehydrogenase subunit B
0.44	306	yzgL	hypothetical protein

Biological network inference

- Goal is to infer (reverse engineer) the topology of a network (e.g., regulatory, signaling), based on direct, indirect or combined association data.
- Identify data patterns that indicate causal influence.
- Networks serve as the basis for dynamical systems modeling and ultimately prediction of cellular responses.

Nodes = Transcription factors
Edges = regulatory events



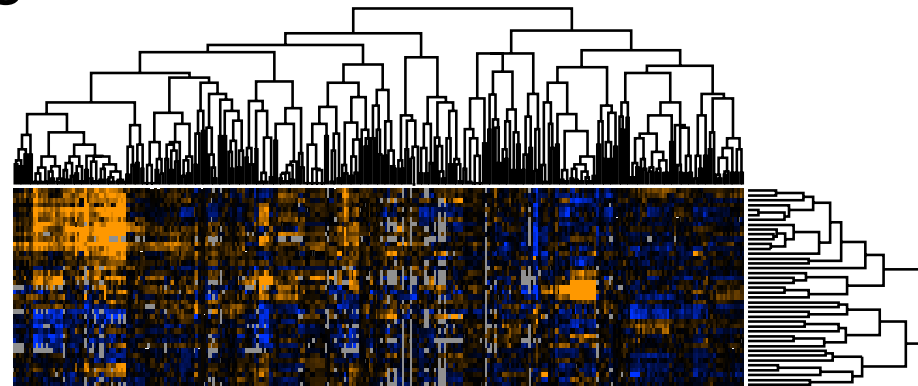
Yeast regulatory network from Maclsaac et al. 2006, based on computational refinement of chromatin immunoprecipitation on chip (ChIP/chip) TF binding site data.

Basic biclustering

A **bicluster** of a data set is a subset of rows that exhibit similar patterns across a subset of columns, or vice versa.

A family of data clustering methods which use a similarity measure (Euclidean distance, Pearson correlation etc.) to compute the input distance matrix for a data clustering algorithm (e.g., hierarchical, k-means). The result are clusters of genes and experiments with similar expression profiles and a 2-D ordering of the data induced by the dendrograms.

Disadvantage: uses all experiments and all genes in comparisons and only 1 bicluster per gene/experiment.



TIGR Multi-experiment viewer, MeV

The next level: bicluster searching and its applications

Family of computationally intensive methods, which search a data matrix for potentially overlapping biclusters.

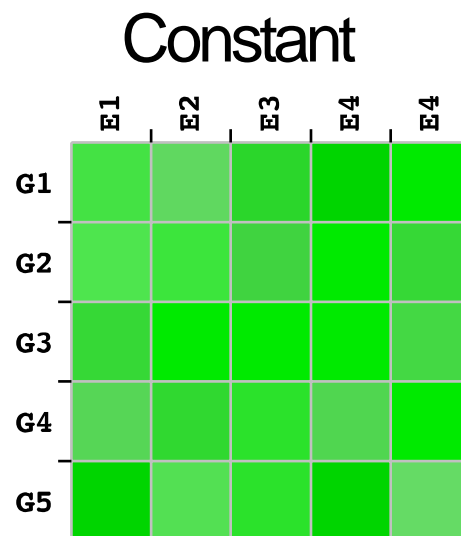
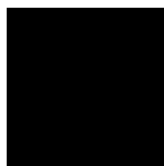
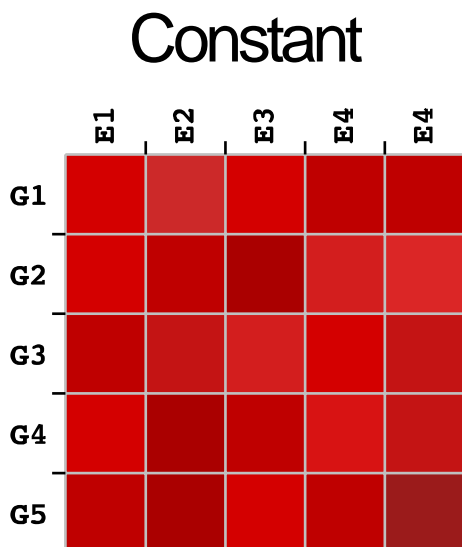
- Discover (overlapping*) sets of genes and experiments exhibiting a (non-binary*) data pattern.
- Test stability of gene+experiment, gene, or experiment sets with respect to a variety* of datasets and statistical criteria.
- Assignment of significance* to patterns in data, removal of noise from results and hypothesis.
- High-throughput experimental data analysis and troubleshooting.

* Increased complexity

Coherent bicluster patterns: biological mechanism signatures

Uniform differential expression:

Common regulation in the form of transcriptional activation or repression.
E.g., genes in an operon, regulon.



Log ratio



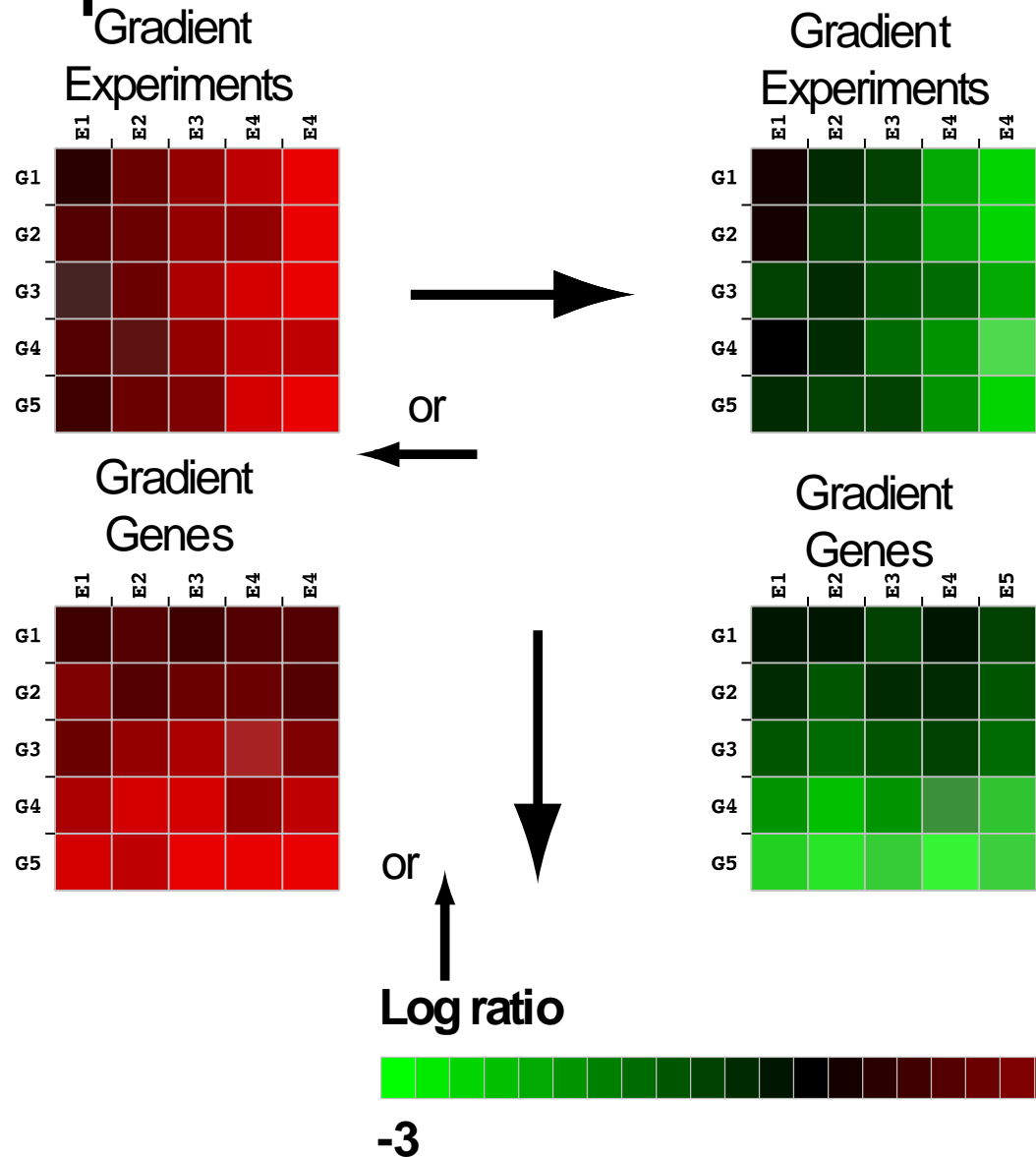
-3

+3

Non-constant coherent bicluster patterns

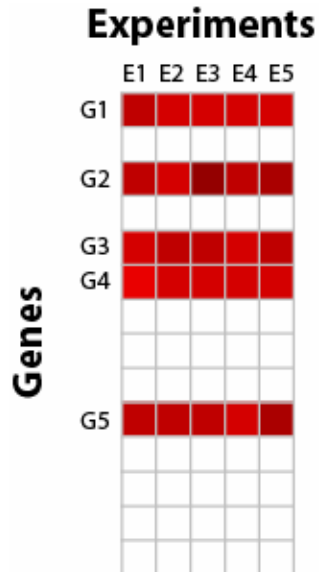
Increasing/decreasing response in time course or condition gradient.

Potentially co-regulated transcripts differing in magnitude of differential expression.

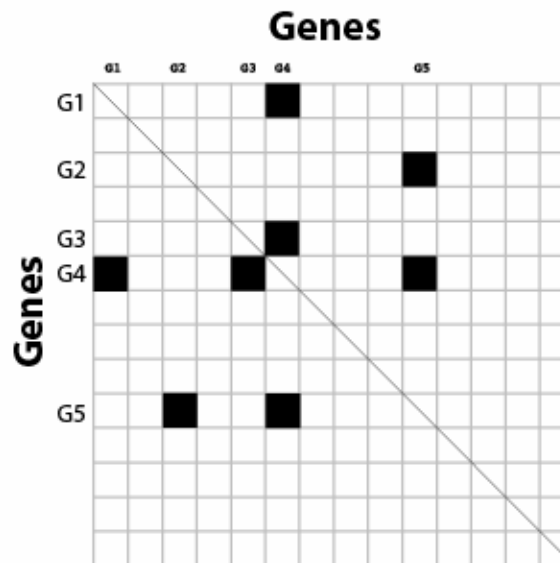


Common functional genomics data types

Gene-by-experiment,
e.g., gene expression



Gene-by-gene, e.g.,
protein interaction matrix



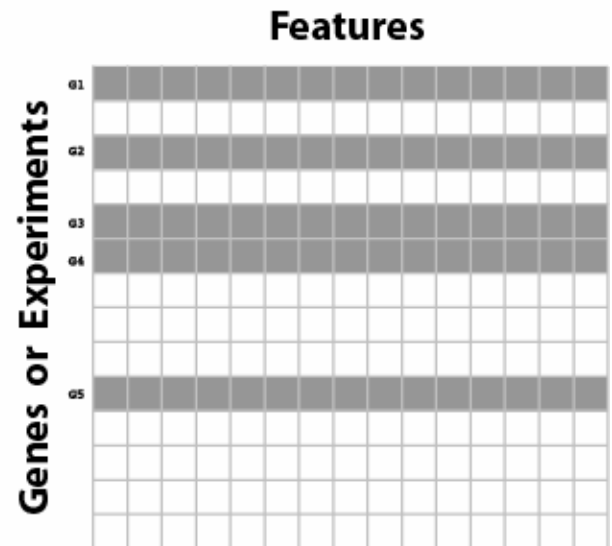
Gene- or experiment- by-features, e.g.:

GO terms

Functional categories

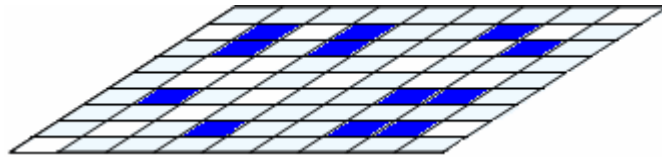
Phylogenetic profiles

Gene, mRNA, and
protein features

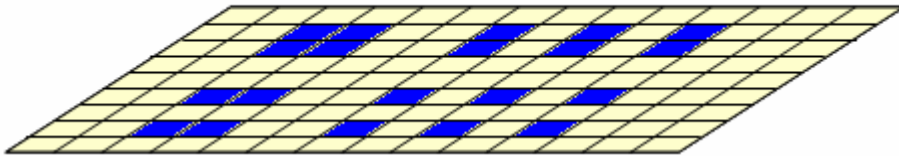


Biclustering as statistical data organization and integration

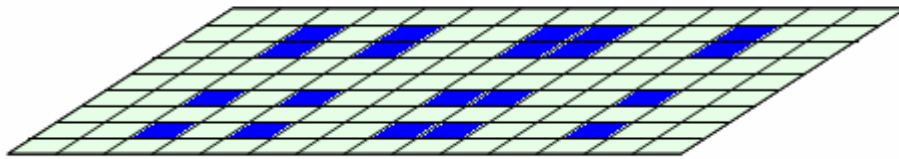
Full criterion =
weighted sum of sub-criteria



Protein by Protein



Gene by Experiment



Gene/Experiment by Feature

Proportion score

$$\text{MSE}(\bar{X}) = E((\bar{X} - \mu)^2) = \left(\frac{\sigma}{\sqrt{n}} \right)^2$$

Row **Mean Squared Error (MSE)**,
other correlation and rank criteria

- Significance score calculated from an empirical null distribution created from random draws of all allowed bicluster sizes.
- Probability that a value as extreme or more extreme would occur if bicluster was randomly sampled.

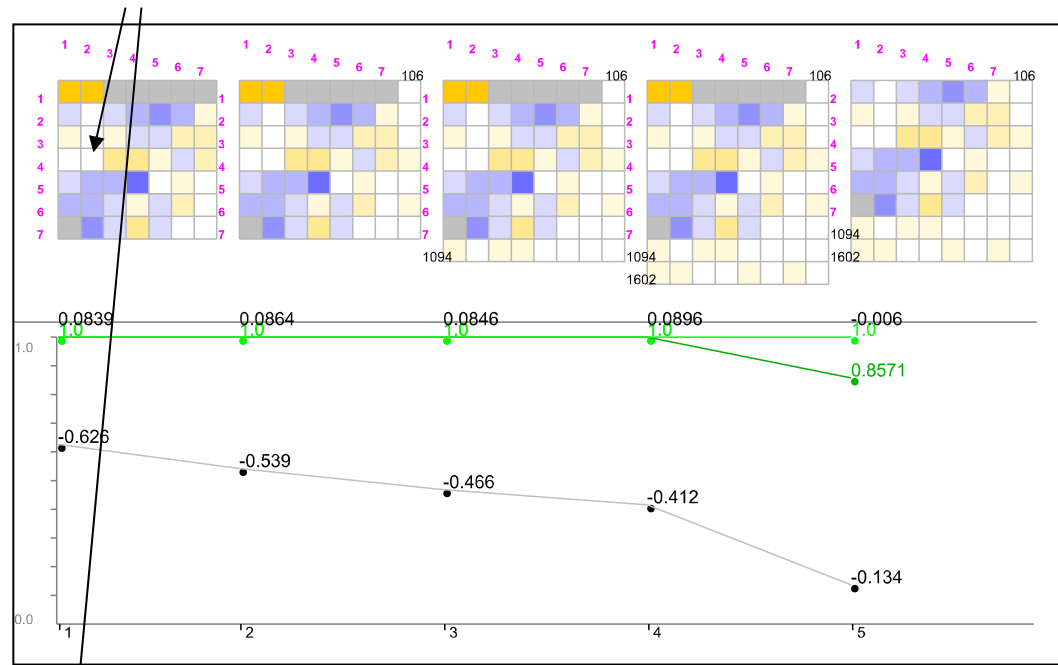
Cross-validated R^2

$$R^2 = \frac{\sum_{i=1}^n (\hat{y}_i - \bar{y})^2}{\sum_{i=1}^n (y_i - \bar{y})^2}$$

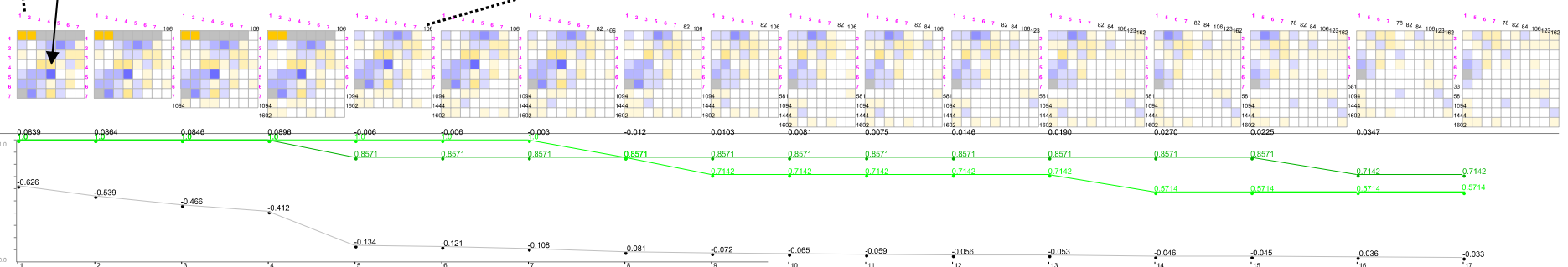
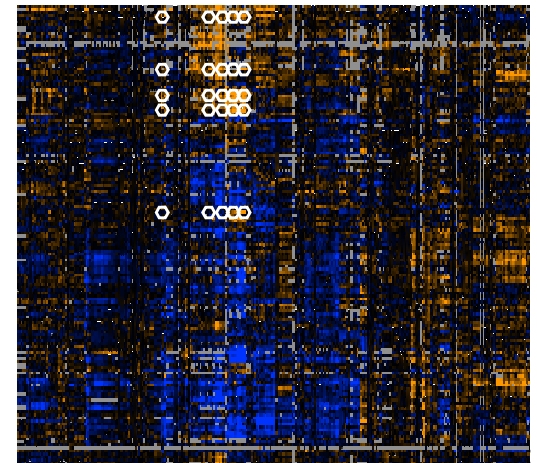
- Calculated using data-adaptive software with polynomial spline fitting.
- Selects subset of features using cross-validation.

Bicluster search trajectory

Random 7x7
starting block



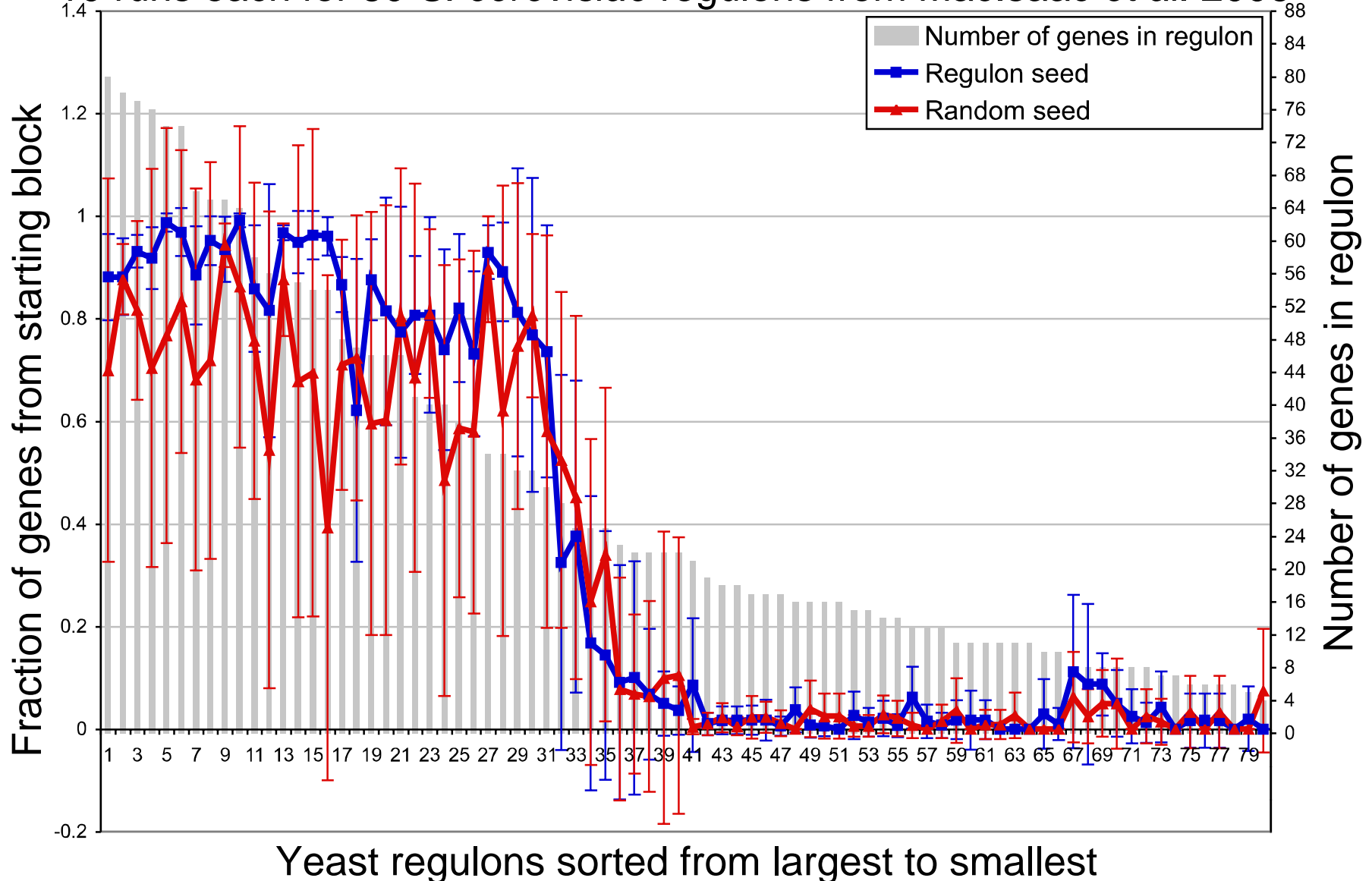
Current bicluster



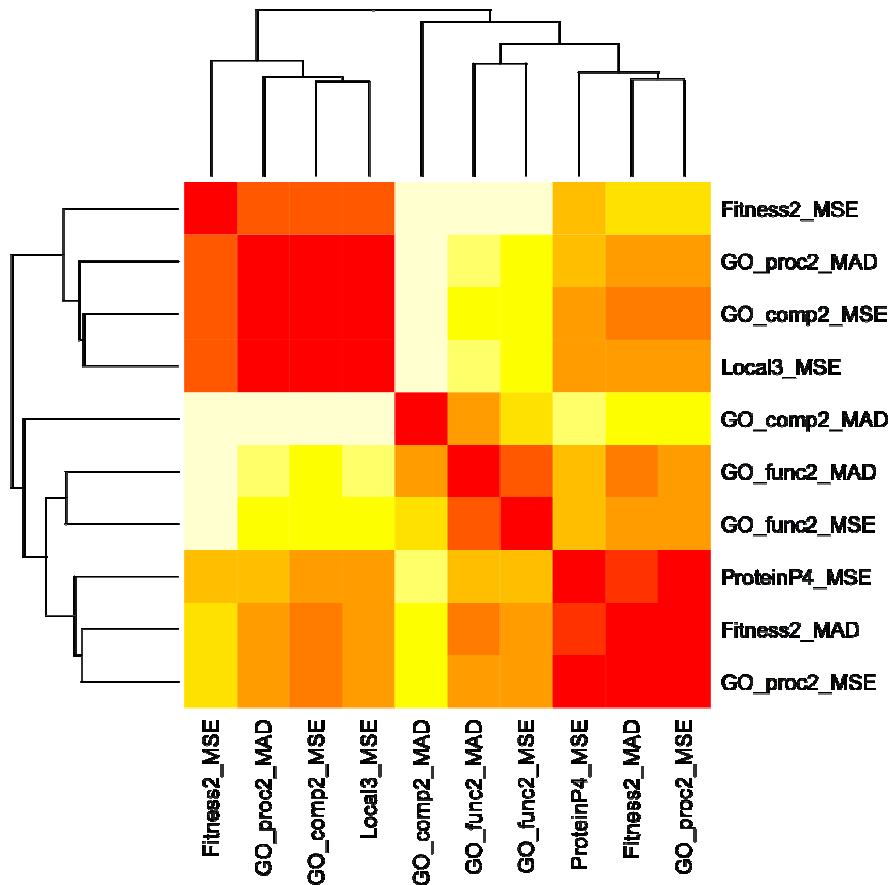
Yeast regulon 'stability'

Results from novel statistical data fusion algorithm:

10 runs each for 80 *S. cerevisiae* regulons from MacIsaac *et al.* 2006



Comparing features and criteria: bicluster membership



Bicluster **A** and **B** overlap:

$$\bigcap \text{Genes}_A, \text{Genes}_B$$

$$\sqrt{\text{Genes}_A \times \text{Genes}_B}$$

$$\bigcap \text{Experiments}_A, \text{Experiments}_B$$

$$\sqrt{\text{Experiments}_A \times \text{Experiments}_B}$$

10 final biclusters

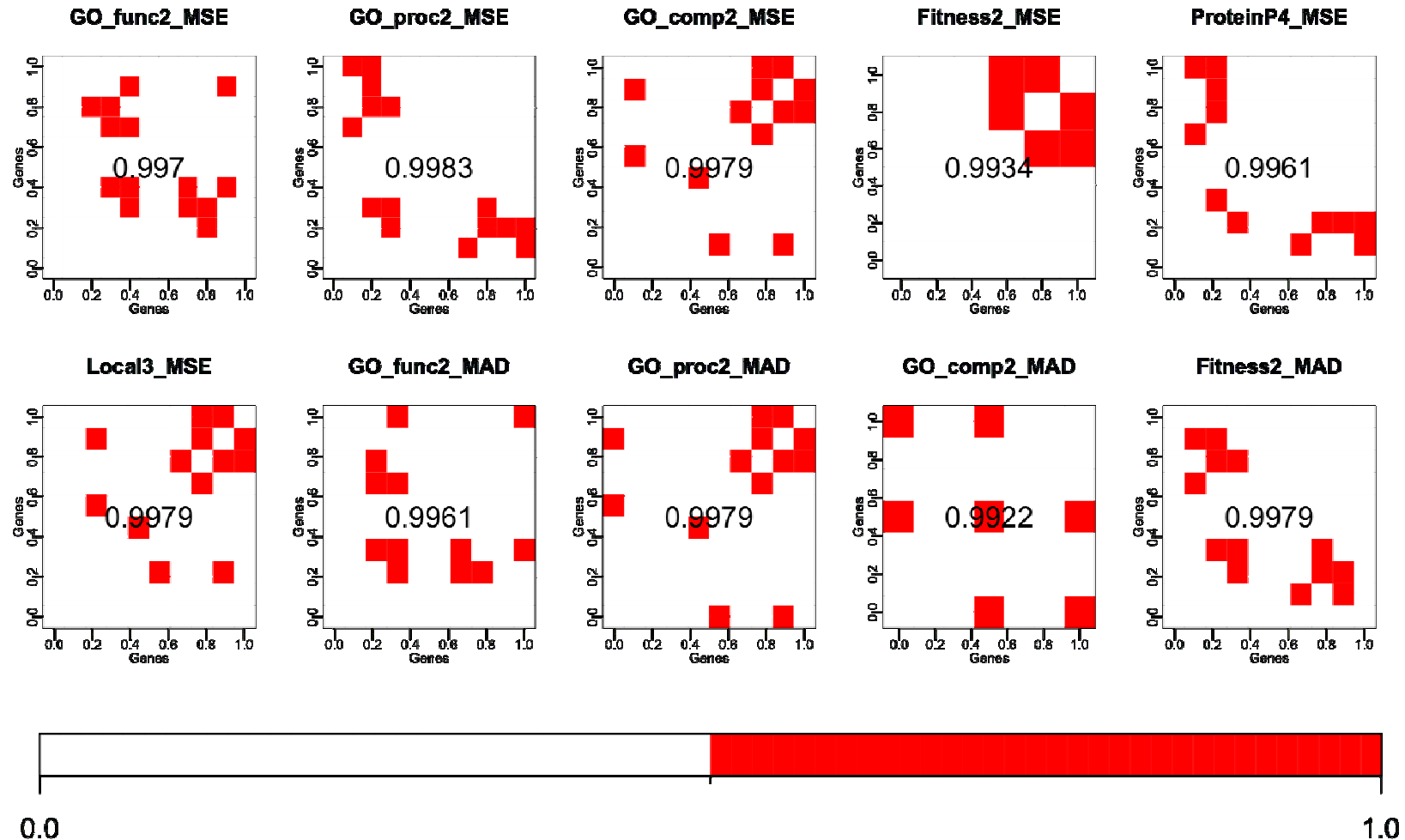
Single set of runs

Same starting bicluster

Varying feature set

Varying criteria

Protein-protein interaction profiles for final biclusters



Perspectives

- Next questions
 - How do different features and datasets contribute to known regulon recovery?
 - What are the properties of known regulons?
- Next additions to algorithm
 - Methods
 - Forward selection
 - Post-analysis toolbox
 - Datasets
 - Sequence motifs
 - Pathways and metabolites
- What are the hallmarks of success?
 - Evaluate recovery of known regulons in presence of noise
 - Discovery of novel regulons
 - Dynamical modeling based on predicted regulons

Acknowledgements

Biclustering data fusion algorithm

Adam Arkin

Mark van der Laan

Cathy Tuglus

PCAP

Swapnil Chabra

John-Marc Chandonia



VIMSS & www.microbesonline.org

From left to right:

Dylan Chivian

Paramvir Dehal

Morgan Price

...

Adam Arkin

Keith Keller

Jason Baumohl (not shown)